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## Kidney disorders and management through herbs: A Review

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### ABSTRACT

Kidneys have a vital role in the normal physiology of humans. Worldwide chronic kidney disease has become a major cause for disability and in worst circumstances leads to death. Major renal disorders occur due to diabetes and its complications termed as diabetic nephropathy (DN). Also nephrolithiasis occurs due to presence of organic debris of carbohydrates, lipids and proteins and supersaturation with calcium oxalate in the renal system. The article comprises of various herbs proven to be used in management of these disorders

**Keywords:** Diabetes, diabetic nephropathy, kidney stone, herbs, chronic kidney disease (CKD).

### INTRODUCTION

The important role of kidneys in normal physiology comprises plasma filtration of metabolic waste products, regulation of plasma volume, hormone secretion and acid-base balance. Any changes in the above indicators lead to a large number of diverse, life threatening renal diseases. Globally, the 12th cause of death in humans is due to chronic kidney disease (CKD) and leads to 17th cause of disability. People with CKD are more prone to cardiovascular disorders (CVD) rather than to reach end-stage renal disease (ESRD) [1]. Around 30% of diabetes mellitus patients (DM) fall ill with diabetic nephropathy (DN) and CKD incidence. According to the Diabetes Atlas 2006 (India), patient's population with DM is presumed to rise to 69.9 million by 2025 in the absence of preventive measures [2]. "Screening and Early Evaluation of Kidney Disease" (SEEK), a voluntary health screening program which is community-based started in 2006 in India performed analysis of urine and serum creatinine of people. SEEK announced high prevalence of CKD approximately 17.4% applying a glomerular filtration (eGFR) formula. Indian CKD Registry states that diabetes (all types) is the cause of kidney disease in 30% of the patients enlisted in their studies. Just 20% of the ESRD registered patients are on some renal replacement therapies (RRT) [3]. The limitation of ESRD is that it is inpatient thus hospital-based and not an exact figure of population suffering from ESRD. The yearly incidences of ESRD in India is approximately 150–200 per million population (pmp) and Diabetes mellitus is an essential cause of CKD in around 30–40% of these patients [4]. It is evaluated that only 10–20% patients in India with ESRD carry out long-term RRT. In India 3,500 new kidney transplant take place annually, about 3,000 new continuous ambulatory peritoneal dialysis (CAPD) gets initiated and 15,000 new maintenance hemodialysis (MHD) patients [5].

Urine microalbuminuria, especially in patients with DM, is a first indicator in patients at risk of kidney disease well ahead the rise in gross proteinuria or elevated serum creatinine. Deviation in the level of GFR measured from serum creatinine indicates any kidney disease at an early stage [6]. Diabetic nephropathy can be explained with change in levels of microalbuminuria, succeeded by macroproteinuria and also reduction in GFR. Moreover, renal disease in DM can happen without excretion of protein in urine of patients with DM and kidney disorders [7]. Plasma filtration and most of tubular reabsorption occurs in renal cortex, an important functional portion of the kidney present in between the renal capsule and renal medulla, comprising of glomeruli, proximal and distal tubules. Among all studies renal pathologies, diabetic nephropathy (DN) is predominantly most common causes of renal insufficiency culminating in renal failure. DN is a generally a glomerular disorder but recent scientific literature have focused on the marked changes in tubulointerstitial parameters which strongly suggest that approaches concentrating only on either glomeruli or tubules are not sufficient for thorough knowledge of the pathophysiology of complicated renal diseases such as DN [8].

CKD is highly prevalent in south Asian population viz. India, Sri Lanka Bangladesh and Pakistan, and black people due to higher rates of occurrence of diabetes in Asians and higher rates of increased blood pressure in Caribbean and African people. The socioeconomic status and ethnic origin of several communities in both higher and lower income countries are reason for their greater risk than others [9].

### DIABETIC NEPHROPATHY

DN is a major complication mainly associated with type 2 diabetes that leads to ESRD. In India, DN is expected to develop in 6.6 million of the 30 million patients suffering from DM by 2030 [10]. DN is one of the major “microvascular” disorder related to diabetes. The renal lesions which develop in type 1 or 2 diabetes mellitus, are similar [11]. DN is characterized by an increase in various things viz. kidney size, urinary albumin excretion, glomerular volume and kidney function chased by the accumulation of glomerular extracellular matrix, glomerular sclerosis and tubular fibrosis. Proteinuria, hypertension, and progressive renal insufficiency indicate last-stage overt DN [12]. Diabetic kidney disease is reported in about 15%–25% of type I diabetes patients and 30%–40% of patients with type II diabetes. The pathophysiology of DN comprises of hyperfiltration and development of microalbumin in urine which is followed by deterioration of kidney functions associated with extracellular and cellular disruption in both places that is glomerular and tubulo-interstitial regions of kidney [13]. It also includes hypertrophy/hyperplasia of glomerulus and the tubules, thickening of tubular basement membranes, thickening of glomerular, and expansion of tubulo-interstitial as well as mesangial compartments [14]. There are changes in hyalinization of arterioles, thickening of branches of intrarenal arteries which causes impairment in autoregulation of glomerular microcirculation, that could ultimately damage the kidney.

### KIDNEY STONE

Renal colic is the first manifestation of renal stone disease. The formation of solid phases in urinary passages is described as “Nephrolithiasis”, whereas the accumulation and aggregation of salts in renal parenchyma is termed as “nephrocalcinosis”. Nephrocalcinosis is very common and can develop or cannot into nephrolithiasis. Formation of kidney stone is a complex process including chronicle events, viz. crystal nucleation, its growth, aggregation, and crystal retention inside the renal tubules [15]. Adequate management of the patient can be done by (i) attenuating the pain; (ii) by favoring progression and spontaneous expulsion of stones from body and (iii) by preventing obstructive and infectious complications. Further consideration should be prevention of new stone formation and in some cases dissolution of stones only after acute episodes of pain have been managed completely. Therapeutic treatment involves use of opioids,

nonsteroidal anti-inflammatory drugs (NSAIDs) and spasmolytics. Efficacy of available drugs is primarily dependent on individual response, though NSAID such as ketorolac, alone or in association with morphine, have given better results to reduce pain. Injections of Voveron/ Diclofenac are administered to patients with acute pain [16]. The chronic mild hyperoxaluria is primary cause of stone formation in humans. All crystals, accumulated salts and stones, which are spontaneously formed in humans, contain organic material viz. carbohydrates, lipids and proteins occluded within as well as on their surfaces for providing architectural integrity, otherwise the stone may crumble and disintegrate in particles. Thus, the interaction between crystals and organic material is very critical and important [15]. According to the chemical composition, kidney stones are classified into various types. As per the literature, calcium oxalate (CaOx) is predominant component of stones accounting for 80% or more of all stones found in kidney. The remaining 20% colics comprise struvite stone, cystine, uric acid, and other types of stones. Crystallization and subsequent lithogenesis happens with many solutes which are found in urine. Urine should be supersaturated with respect to the colic base material for formation into bigger crystals. Thus, lowering supersaturation is effective for preventing stone recurrence. Larger than 5 mm stones fail to pass through urine and need interventional by techniques such as ureteroscopy (URS), percutaneous nephrolithotomy (PNL) or extracorporeal shock wave lithotripsy (ESWL), for their removal from the body. As a result of excessive protein intake with CaOx stones, urine uric acid excretion generally elevates. Hyperuricosuria reduces the solubility of calcium oxalate. It encourages formation of stone by heterologous nucleation on the surface of monosodium urate levels. One approach to avoid kidney stone formation is to stop retention of crystals. Reactive oxygen species (ROS) appear to be responsible for injury to renal cells, therefore a reduction in tissue oxidative stress could also be an effective therapeutic measure for recovery [17].

Thus, interdisciplinary research between pharmacologist, pharmacognosists and clinical investigators is essential to develop new plant-derived high potency, high-quality natural products to prevent or completely treat DN and renal colics.

Dialysis and transplantation are the methods of management of ESRD. In developing countries, ethno-medicinal plants have traditionally been used for the treatment of diabetes as well as related complications. In fact, recent pre-clinical and clinical studies have confirmed beneficial effects of many plants on some or the other processes connected with reduced kidney functions in experimental animals [18, 19]. Some of the active phytochemicals are responsible for their potential activities. The therapeutic effect and pharmacological properties of few ethno-botanical herbs which have been used traditionally in the management of DN and urolithiasis have been established [Table 1].

**Table 1:** Plants used in treatment of kidney disorders

Cisplatin induced nephrotoxicity				
Plant Name	Common Name	Family	Plant part/ extract	Reference
<i>Aerva Javanica</i>	Dessert cotton	Amaranthaceae	Fresh roots	[20]
<i>Aerva lanata</i>	Mountain knotgrass	Amaranthaceae	Aerial plants	[21]
<i>Aesculus hippocatanum</i>	Horse chestnut	Sapindaceae	Seeds	[22]
<i>Aloe barbadensis</i>	Aloe vera	Asphodelaceae	Leaf	[23]
<i>Bauhinia variegata</i>	Kachnar	Caesalpiniaceae	Stems	[24]
<i>Carica papaya</i>	Papaya	Caricaceae	Seeds	[25]
<i>Cassia auriculata</i>	Matara tea/ Tarwar	Fabaceae	Roots	[26]
<i>Ceratonia siliqua</i>	Carob	Fabaceae	Pods and leaves	[27]
<i>Crataeva nurvala</i>	Varuna	Capparaceae	Stem bark	[28]
<i>Cucurbita pepo</i>	Pumpkin	Cucurbitaceae	Fruits	[25]
<i>Cyclea peltata</i>	Rajpatha	Menispermaceae	Leaves	[29]

<i>Dichrostachys cinera</i>	Chinese lantern	Mimosaceae	Roots	[30]
<i>Ficus religiosa</i>	Peepal	Moraceae	Latex	[31]
<i>Kigelia africana</i>	Sausage	Bignoniaceae	Matured fruits	[32]
<i>Lepidium sativum</i>	Pepper Wort	Cruciferae	Seeds	[33]
<i>Pedaliium murex</i>	Bada gokhru	Pedaliaceae	Fruits	[34]
<i>Picrorhiza kurroa</i>	Kutki	Scrophulariaceae	Rhizome	[35]
<i>Silybum marianum</i>	Milk thistle	Asteraceae	Seeds	[36]
<i>Veronia cinerea</i>	Sahadevi	Compositae	Aerial parts	[37]
<b>Gentamicin induced nephrotoxicity</b>				
<i>Aloe barbadensis</i>	Aloe vera	Asphodelaceae	Leaves	[23]
<i>Aegle marmelos</i>	Wood apple	Rutaceae	Leaves	[38]
<i>Aerva lanata</i>	Mountain knotgrass	Amaranthaceae	Whole plant	[21]
<i>Orthosiphon stamineus</i>	Kidney tea	Lamiaceae	Whole plant	[39]
<i>Strychnos potatorum</i>	Clearing nut	Loganiaceae	Seeds	[40]
<b>Diabetic nephropathy</b>				
<i>Allium sativum</i>	Garlic	Alliaceae	Cloves	[41]
<i>Andrographis paniculata</i>	Kalmegh	Acanthaceae	Roots	[42]
<i>Astragalus membranaceus</i>	Milk vetch	Fabaceae	Roots	[43]
<i>Berberis integerrima</i>	Barberry	Berberidaceae	Roots	[44]
<i>Brassica oleracea</i>	Red Cabbage	Brassicaceae	Leaves	[45]
<i>Camellia sinensis</i>	Green tea	Theaceae	Leaves	[46]
<i>Cinnamomum zeylanicum</i>	Dalchini	Lauraceae	Oil	[47]
<i>Curcuma longa</i>	Turmeric	Zingiberaceae	Curcumin	[48]
<i>Ekebergia capensis</i>	Cape ash	Meliaceae	Leaves	[49]
<i>Eugenia jambolana</i>	Black berry	Myrtaceae	Seeds	[50]
<i>Ficus thonningii</i>	Blume	Moraceae	Stem bark	[51, 52]
<i>Foeniculum vulgare</i>	Fennel	Apiaceae	Fruits	[53]
<i>Fragaria × ananassa</i>	Strawberry	Rosaceae	Leaves	[54]
<i>Ganoderma lucidum</i>	Mushroom	Polyporaceae	Fruits	[55]
<i>Ginkgo biloba</i>	Maiden Hair	Gikgoaceae	Leaves	[56]
<i>Glycyrrhiza uralensis</i>	Chinese liquorice	Fabaceae	Roots	[57]
<i>Gongronema latifolium</i>	Amaranth globe	Asclepiadaceae	Leaves	[58]
<i>Gymnema montanum</i>	Bidaria Tingens Deche	Asclepidaceae	Leaves	[59]
<i>Helianthus annuus</i>	Sunflower	Asteraceae	Leaves	[60]
<i>Helichrysum ceres</i>	Beentje	Asteraceae	Leaves	[61]
<i>Hypoxis hemerocallidea</i>	African potato	Hypoxidaceae	Corm	[62, 63]
<i>Indigofera tinctoria</i>	True indigo	Fabaceae	Leaves	[64]
<i>Linum usitatissimum</i>	Flax seeds	Linaceae	Seeds	[65]
<i>Momordica charantia</i>	Bitter gourd	Cucurbitaceae	Seeds	[50]
<i>Moringa oleifera</i>	Drumstick/Horseradish	Moringaceae	Leaves	[66]
<i>Olea europaea</i>	European olive	Oleaceae	Leaves	[67, 68]
<i>Opuntia megacantha</i>	Prickly pear	Cactaceae	Leaves	[69]
<i>Panax quinquefolius</i>	American ginseng	Araliaceae	Roots	[70]
<i>Persea americana</i>	Avacado	Lauraceae	Leaves	[71]
<i>Pterocarpus santalinus</i>	Red sandal wood	Fabaceae	Heartwood	[72]
<i>Rheum officinale</i>	Rhubarb	Polygonaceae	Resin	[73]
<i>Salvia miltiorrhiza</i>	Chinese sage	Lamiaceae	Roots	[74]
<i>Sclerocarya birrea</i>	Marula	Anachardiaceae	Stem bark	[75]
<i>Sesamum indicum</i>	Sesame	Pedaliaceae	Seeds	[76]
<i>Silybum marianum</i>	Milk thistle	Asteraceae	Seeds	[77, 78]
<i>Tectona grandis</i>	Teak	Lamiaceae	Bark	[79]
<i>Terminalia chebula</i>	Black myrobalan	Combretaceae	Fruits	[80]

<i>Vitis vinifera</i>	Grape vine	Vitaceae	Fruit wine	[81]
<i>Zingiber officinale</i>	Ginger	Zingiberaceae	Rhizome	[41]
<b>Nephrolithiasis</b>				
<i>Aerva lanata</i>	Chaya/ Mountain knotgrass	Amaranthaceae	Leaves	[82, 83]
<i>Ammannia baccifera</i>	Monarch redstem	Lythraceae	Aerial parts	[84]
<i>Asparagus racemosus</i>	Shatavari	Asparagaceae	Whole plant	[85]
<i>Alisma orientale</i>	Takusha	Alismataceae	Rhizome	[86]
<i>Bergenia ciliata</i>	Megaseas	Saxifragaceae	Leaves	[87]
<i>Boerhaavia diffusa</i>	Punarnava	Nyctaginaceae	Roots	[88]
<i>Bryophyllum pinnatum</i>	Pattharchata	Crassulaceae	Leaves	[88]
<i>Citrus limon</i>	Lemon	Rutaceae	Lemon juice	[89, 90]
<i>Costus spiralis</i>	Cana-do-brejo	Zingiberaceae	Whole plant	[91]
<i>Crataeva nurvala</i>	Varuna	Capparaceae	Bark	[92]
<i>Cyclea peltata</i>	Rajpatha	Menispermaceae	Roots	[93]
<i>Cynodon dactylon</i>	Bermuda grass	Poaceae	Fresh plant	[94]
<i>Daucus carota</i>	Carrot	Apiaceae	Root	[88]
<i>Desmodium styracifolium</i>	Coin-leaf desmodium	Leguminosae	Triterpene	[95]
<i>Helianthus annuus</i>	Sunflower	Asteraceae	Leaves	[60]
<i>Herniaria hirsuta</i>	Hairy rupturewort	Caryophyllaceae	Fresh herb	[96]
<i>Hibiscus sabdariffa</i>	Roselle	Malvaceae	Leaves	[97]
<i>Ipomoea eriocarpa</i>	Tiny morning glory	Convolvulaceae	Leaves	[98]
<i>Jasminum auriculatum</i>	Jasmine	Oleaceae	Flowers	[99]
<i>Mimosa pudica</i>	Laajvanti	Mimosaceae	Leaves	[100]
<i>Moringa oleifera</i>	Drumstick/ Horseradish	Moringaceae	Root	[101]
<i>Musa sapientum</i>	Banana	Musaceae	Stem	[102]
<i>Nigella sativa</i>	Black caraway/ kalonji	Ranunculaceae	Seeds	[103]
<i>Orthosiphon stamineus</i>	Java tea	Lamiaceae	Leaves	[104]
<i>Phyllanthus niruri</i>	Gale of the wind	Phyllanthaceae	Whole plant	[105]
<i>Punica granatum</i>	Anaar	Lythraceae	Fruit juice	[106]
<i>Quercus stenophylla</i>	Oak	Fagaceae	Leaves	[107]
<i>Randia echinocarpa</i>	Chacua	Rubiaceae	Fruit	[108]
<i>Raphanus sativus var. nigra</i>	Radish	Cruciferae	Tubercle	[109]
<i>Rosa canina</i>	Dog rose	Rosaceae	Fruit	[110]
<i>Rotula aquatic</i>	Pashanbhed	Boraginaceae	Extract	[111]
<i>Rubia cordifolia</i>	Indian madder	Rubiaceae	Roots	[112]
<i>Rubia tinctorium</i>	Dyer's madder	Rubiaceae	Roots	[113]
<i>Sesbania grandiflora</i>	Agastya	Fabaceae	Leaves	[114]
<i>Spergularia purpurea</i>	Purple sandspurry	Caryophyllaceae	Whole plant	[115]
<i>Tribulus terrestris</i>	Chhota gokhru	Zygophyllaceae	Fruits, roots	[116]
<i>Trigonella foenum graecum</i>	Methi	Fabaceae	Seeds	[117]
<i>Zea mays</i>	Maize	Poaceae	Styles	[118]

## CONCLUSION

The review summarizes all the plants which have been scientifically proven for the treatment of DN and urolithiasis and thus a systematic research attempt is need of the hour to explore botanicals as alternative and/or complementary medicines which could be formulated in potent dosage forms so as to be easily available to people all around the world.

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## REFERENCES

1. Ritz E, Rychlik I, Locatelli F, Halimi S. End-stage renal failure in type 2 diabetes: A medical catastrophe of worldwide dimensions. *Am. J. Kidney Dis.* 1999; 34(5):795-808.
2. Sicree R, Shaw J, Zimmet P. Diabetes and impaired glucose tolerance. In: Gan D, editor. *Diabetes Atlas*. 3rd ed. Brussels: International Diabetes Federation, 2006; 15-109.
3. Anonymous. CKD registry of India: Indian Society of Nephrology, 2017. Available from <http://www.ckdri.org>. (2 Jan. 2017)

4. Grassmann A, Gioberge S, Moeller S, Brown G. ESRD patients in 2004: Global overview of patient numbers, treatment modalities and associated trends. *Nephrol. Dial. Transplant.* 2004; 20(12):2587-2593.
5. Agarwal SK, Srivastava RK. Chronic kidney disease in India: Challenges and solutions. *Nephron Clin. Pract.* 2009; 111(3):c197-c203.
6. Kalantar-Zadeh K, Amin AN. Toward more accurate detection and risk stratification of chronic kidney disease. *JAMA* 2012; 307(18):1976-1977.
7. Schwenger V, Mussig C, Hergesell O, Zeier M, Ritz E. Incidence and clinical characteristics of renal insufficiency in diabetic patients. *Dtsch. Med. Wochenschr.* 2001; 126(47):1322-1326.
8. Zhao Y, Denner L, Haidacher SJ, LeJeune WS, Tilton RG. Comprehensive analysis of the mouse renal cortex using two-dimensional HPLC--tandem mass spectrometry. *Proteome Sci.* 2008; 6(15):1-15.
9. Gupta MC, Trilok C. A critical review on commonly used herbal drugs in CKD. *J. Med. Plant Stud.* 2015; 3(4):44-47.
10. Baig RM, Gillani WS, Sulaiman SA, Krishna RD, Narayanan K. Epidemiology of diabetic nephropathy in the poor patients from rural south-east India. *Int. J. Nutr. Food Sci.* 2011; 4(1):53-61.
11. Fioretto P, Mauer M. Histopathology of diabetic nephropathy. *Semin. Nephrol.* 2007; 27(2):195-207.
12. Sheela N, Jose MA, Sathyamurthy D, Kumar BN. Effect of silymarin on streptozotocin-nicotinamide-induced Type 2 diabetic nephropathy in rats. *Iran. J. Kidney Dis.* 2013; 7(2):117-123.
13. Kanwar YS, Wada J, Sun L, Xie P, Wallner EI, Chen S, *et al.* Diabetic nephropathy: Mechanisms of renal disease progression. *Exp. Biol. Med.* 2008; 233(1):4-11.
14. Mason RM, Wahab NA. Extracellular matrix metabolism in diabetic nephropathy. *J. Am. Soc. Nephrol.* 2003; 14(5):1358-1373.
15. Khan SR. Animal models of kidney stone formation: An analysis. *World J. Urol.* 1997; 15(4):236-243.
16. Marangella M, Vitale C, Bagnis C, Petrarulo M, Tricerri A. Use of drugs for nephrolithiasis. *Clin. Cases Miner. Bone Metab.* 2008; 5(2):131-134.
17. Butterweek V, Khan SR. Herbal Medicines in the Management of Urolithiasis: Alternative or Complementary? *Planta Med.* 2009; 75(10):1095-1103.
18. Wang XM, Guan SH, Liu RX, Sun JH, Liang Y, Yang M, *et al.* HPLC determination of four triterpenoids in rat urine after oral administration of total triterpenoids from *Ganoderma lucidum*. *J. Pharm. Biomed. Anal.* 2007; 43(3):1185-1190.
19. Nakagawa T, Goto H, Hikiami H, Yokozawa T, Shibahara N, Shimada Y. Protective effects of keishibukuryogan on the kidney of spontaneously diabetic WBN/Kob rats. *J. Ethnopharmacol.* 2007; 110(2):311-317.
20. Movaliyaa V, Khamarb D, Setty MM. Nephroprotective activity of aqueous extract of *Aerva Javanica* roots in cisplatin-induced renal toxicity in rats. *Pharmacologyonline*. 2011; 1:68-74.
21. Shirwaikar A, Issac D, Malini S. Effect of *Aerva lanata* on cisplatin and gentamicin models of acute renal failure. *J. Ethnopharmacol.* 2004; 90(1):81-86.
22. Elmas O, Erbas O, Yigiturk G. The efficacy of *Aesculus hippocastanum* seeds on diabetic nephropathy in a streptozotocin-induced diabetic rat model. *Biomed. Pharmacother.* 2016; 83:392-396.
23. Chatterjee P, Mukherjee A, Nandy S. Protective effects of the aqueous leaf extract of *Aloe barbadensis* on gentamicin and cisplatin-induced nephrotoxic rats. *Asian Pac. J. Trop. Biomed.* 2012; 9(1):S1754-S1763.
24. Pani SR, Mishra S, Sahoo S, Panda PK. Nephroprotective effect of *Bauhinia variegata* (Linn.) whole stem extract against cisplatin-induced nephropathy in rats. *Indian J. Pharmacol.* 2011; 43(2):200-202.
25. Debnath S, Babre N, Manjunath YS, Mallareddy V, Parameshwar P, Hariprasath K. Nephroprotective evaluation of ethanolic extract of the seeds of papaya and pumpkin fruit in cisplatin-induced nephrotoxicity. *J. Pharm. Sci. Technol.* 2010; 2(6):241-246.
26. Shirwaikar A, Rajagopal PL, Malini S. Effect of *Cassia auriculata* Linn. root extract on cisplatin and gentamicin-induced renal injury. *Phytomedicine* 2005; 12(8):555-560.
27. Ahmed MM. Biochemical studies on nephroprotective effect of *Carob* (*Ceratonia siliqua* L.) growing in Egypt. *Nat. Sci.* 2010; 8(3):41-47.
28. Shelke TT, Bhaskarb VH, Adkara PP, Jhaa U, Oswala RJ. Nephroprotective activity of ethanolic extract of stem barks of *Crataeva nurvala* Buch Hum. *Int. J. Pharm. Sci. Res.* 2011; 2(10):2712-2717.
29. Vijayan FP, Rani VK, Vineesh VR, Sudha KS, Michael MM, Padikkala J. Protective effect of *Cyclea peltata* Lam on cisplatin-induced nephrotoxicity and oxidative damage. *J. Basic Clin. Physiol. Pharmacol.* 2007; 18(2):101-114.
30. Adikay S, Koganti B, Prasad KVSRRG. Effect of alcoholic extract of root of *Dichrostachy scinerea* Wight and Arn. against cisplatin-induced nephrotoxicity in rats. *Nat. Prod. Rad.* 2009; 8(1):12-18.
31. Yadav YC, Srivastava DN, Saini V, Singhal S, Seth AK, Kumar S, *et al.* Nephroprotective and curative activity of methanolic extract of *Ficus religiosa* L. latex in Albino rats using cisplatin-induced nephrotoxicity. *Pharmacologyonline* 2011; 1:132-139.
32. Azu OO, Francis IOD, Abraham AO, Crescie CN, Stephen OE, Abayomi OO. Protective agent, *Kigelia africana* fruit extract, against cisplatin-induced kidney oxidant injury in Sprague--Dawley rats. *Asian J. Pharm. Clin. Res.* 2010; 3(2):84-88.
33. Yadav YC, Srivastav DN, Seth AK, Saini V, Yadav KS. Nephroprotective activity of ethanolic extract *Lepidium sativum* L. seeds in albino rats using cisplatin-induced acute renal failure. *Int. J. Pharm. Sci. Res.* 2010; 4(3):64-68.
34. Rajashekar V, Rao EU, Srinivas P. Biological activities and medicinal properties of *Gokhru* (*Pedalium murex* L.). *Asian Pac. J. Trop. Biomed.* 2012; 2(7):581-585.
35. Gadgoli CH, Yamgar S, Sali L, Salkar R, Jain NK. Studies on nephroprotective and nephrocurative activity of ethanolic extract of *Picrorhiza kurroa* Royle and *Arogyawardhini bati* in rats. *Int. J. Pharm. Technol.* 2010; 2(3):472-489.
36. Momeni A, Hajigholami A, Geshnizjani S, Kheiri S. Effect of silymarin in the prevention of cisplatin nephrotoxicity, a clinical trial study. *J. Clin. Diagn. Res.* 2015; 9(4):OC11-OC13.
37. Sreedevi A, Bharathi K, Prasad KVSRRG. Effect of *Vernonia cinerea* aerial parts against cisplatin-induced nephrotoxicity in rats. *Pharmacologyonline* 2011; 2:548-555.
38. Kore KJ, Shete RV, Jadhav PJ. Nephroprotective role of *A. marmelos* extract. *Int. J. Res. Pharm. Chem.* 2011; 1(3):617-623.
39. Kannappan N, Madhukar A, Mariymmal, Sindhura PU, Mannavalan R. Evaluation of nephroprotective activity of *Orthosiphon stamineus* Benth extract using rat model. *Int. J. PharmTech Res.* 2010; 2(1):209-215.
40. Varghese R, Moideen MM, Suhail MJM, Dhanapal CK. Nephroprotective effect of ethanolic extract of *Strychnos potatorum* seeds in rat models. *Res. J. Pharm. Biol. Chem.* 2011; 2(3):521-529.
41. Al-Qattan K, Thomson M, Ali M. Garlic (*Allium sativum*) and ginger (*Zingiber officinale*) attenuate structural nephropathy progression in streptozotocin-induced diabetic rats. *Eur. E. J. Clin. Nutr. Metab.* 2008; 3:e62-e71.
42. Rao NK. Anti-Hyperglycemic and renal protective activities of *Andrographis paniculata* roots chloroform extract. *Iran. J. Pharmacol. Therap.* 2006; 5(1):47-50.
43. Li M, Wang W, Xue J, Gu Y, Lin S. Meta-analysis of the clinical value of *Astragalus membranaceus* in diabetic nephropathy. *J. Ethnopharmacol.* 2011; 133(2): 412-419.
44. Ashraf H, Heidari R, Nejadi V, Ilkhanipoor M. Aqueous extract of *Berberis integerrima* root improves renal dysfunction in streptozotocin-induced diabetic rats. *Avicenna J. Phytomed.* 2013; 3(1):82-90.
45. Singh J, Upadhyay AK, Bahadur A, Singh B, Singh KP, Rai M. Anti-oxidant phytochemicals in cabbage (*Brassica oleracea* L. var. capitata). *Sci. Hortic.* 2006; 108(3):233-237.
46. Ribaldo PD, Souza DS, Biswas SK, Block K, Lopes de Faria JM, Lopes de Faria JB. Green tea (*Camellia sinesis*) attenuates nephropathy by down regulating NOX4 NADPH oxidase in diabetic spontaneously hypertensive rats. *J. Nutr.* 2009; 139(1):96-100.
47. Mishra A, Bhatti R, Singh A, Singh Ishar MP. Ameliorative effect of the cinnamon oil from *Cinnamom zeylanicum* upon early stage diabetic nephropathy. *Planta Med.* 2010; 76(5):412-417.
48. Sharma S, Kulkarni SK, Chopra K. Curcumin, the active principle of turmeric (*Curcuma longa*), ameliorates diabetic nephropathy in rats. *Clin. Exp. Pharmacol. Physiol.* 2006; 33(10):940-945.
49. Musabayane CT. The effects of medicinal plants on renal function and blood pressure in Diabetes mellitus. *Cardiovasc. J. Afr.* 2012; 23(8):462-468.
50. Grover JK, Vats V, Rathi SS, Dawar R. Traditional Indian anti-diabetic plants attenuate progression of renal damage in streptozotocin induced diabetic mice. *J. Ethnopharmacol.* 2001; 76(3):233-238.
51. Musabayane CT, Gondwe M, Kamadyaapa DR, Chuturgoon AA, Ojewole JAO. Effects of *Ficus thonningii* (Blume) [Moraceae] stem bark ethanolic extract on blood glucose, cardiovascular and kidney functions of rats, and on kidney cell lines of the proximal (LLC-PK1) and distal tubules (MDBK). *Renal Failure* 2007; 29(4):389-397.
52. Usman H, Abdulrahman F, Usman A. Qualitative phytochemical screening and in vitro antimicrobial effects of methanol stem bark extract of *Ficus thonningii* (Moraceae). *Afr. J. Tradit. Complem. Altern. Med.* 2009; 6(3):289-295.
53. El-Hilaly J, Hmammouchib M, Lyoussi B. Ethnobotanical studies and economic evaluation of medicinal plants in Taounate province (Northern Morocco). *J. Ethnopharmacol.* 2003; 86(2-3):149-158.
54. Ibrahim DS, El-Maksoud MA. Effect of strawberry (*Fragaria × ananassa*) leaves extract on diabetic nephropathy in rats. *Int. J. Exp. Pathol.* 2015; 96(2):87-93.
55. He CY, Li WO, Guo SX, Lin SQ, Lin ZB. Effect of polysaccharides from *Ganoderma lucidum* on streptozotocin-induced diabetic nephropathy in mice. *J. Asian Nat. Prod. Res.* 2006; 8(8):705-711.

56. Zhu HW, Shi ZF, Chen YY. Effect of extract of Ginkgo biloba leaf on early diabetic nephropathy. *Zhongguo Zhong Xi Yi Jie He Za Zhi* 2005; 25(10):889-891.
57. Li J, Lee YS, Choi JS, Sung HY, Kim JK, Lim SS, *et al.* Roasted licorice extracts dampen high glucose-induced mesangial hyperplasia and matrix deposition through blocking Akt activation and TGF beta signaling. *Phytomed.* 2010; 17(10):800-810.
58. Ugochukwu NH, Cobourne MK. Modification of renal oxidative stress and lipid peroxidation in streptozotocin-induced diabetic rats treated with extracts from *Gongronema latifolium* leaves. *Clin. Chim. Acta* 2003; 336(1-2):73-81.
59. Ramkumar KM, Ponmanickam P, Velayuthaprabhu S, Archunan G, Rajaguru P. Protective effect of *Gymnema montanum* against renal damage in experimental diabetic rats. *Food Chem. Toxicol.* 2009; 47(10):2516-2521.
60. Khan NI, Shinge JS, Naikwade NS. Antilithiatic effect of *Helianthus annuus* Linn. leaf extract in ethylene glycol and ammonium chloride induced nephrolithiasis. *Int. J. Pharm. Pharm. Sci.* 2010; 2(4):180-184.
61. Musabayane CT, Munjeri O, Mdege ND. Effects of *Helichrysum ceres* extracts on renal function and blood pressure in the rat. *Renal Failure* 2003; 25(1):5-14.
62. Musabayane CT, Xozwa K, Ojewole JAO. Effects of Hypoxis hemerocallidea (Fisch. & C.A. Mey) [Hypoxidaceae] corm (African Potato) aqueous extract on renal electrolyte and fluid handling in the rat. *Renal Failure* 2005; 27(5):763-770.
63. Ojewole JAO. Antinociceptive, anti-inflammatory and antidiabetic properties of Hypoxis hemerocallidea Fisch. & C.A. Mey. (Hypoxidaceae) corm [African potato] aqueous extract in mice and rats. *J. Ethnopharmacol.* 2006; 103(1):126-134.
64. Bangar AV, Saralay MG. Anti-hyperglycaemic activity of ethanol extract and chloroform extract of *Indigofera tinctoria* leaves in streptozotocin induced diabetic mice (Family-Papilionaceae). *Res. J. Pharm. Biol. Chem. Sci.* 2011; 2(1):444-455.
65. Kaur N, Kishore L, Singh R. Therapeutic effect of *Linum usitatissimum* L. in STZ-nicotinamide induced diabetic nephropathy via inhibition of AGE's and oxidative stress. *J. Food Sci. Technol.* 2017; 54(2):408-421.
66. Omodanisi EI, Aboua YG, Oguntibeju OO. Assessment of the anti-hyperglycaemic, anti-inflammatory and antioxidant activities of the methanol extract of *Moringa oleifera* in diabetes-induced nephrotoxic male Wistar rats. *Molecules* 2017; 22(4):439.
67. Benavente-Garcia O, Castillo J, Lorente J, Ortuno A, Del Rio JA. Antioxidant activity of phenolics extracted from *Olea europaea* L. leaves. *Food Chem.* 2000; 68(4):457-462.
68. Somova LI, Shode FO, Ramnanan P, Nadar A. Antihypertensive, antiatherosclerotic and antioxidant activity of triterpenoids isolated from *Olea europaea*, subspecies *africana* leaves. *J. Ethnopharmacol.* 2003; 84(2-3):299-305.
69. Bwititi P, Musabayane CT, Nhachi CFB. Effects of *Opuntia megacantha* on blood glucose and kidney function in streptozotocin diabetic rats. *J. Ethnopharmacol.* 2000; 69(3):247-252.
70. Kim HY, Kang KS, Yamabe N, Nagai R, Yokozawa T. Protective effect of heat-processed American ginseng against diabetic renal damage in rats. *J. Agric. Food Chem.* 2007; 55(21):8491-8497.
71. Afzal M, Khan NA, Ghufuran A, Iqbal A, Inamuddin M. Diuretic and nephroprotective effect of jawarish zarooni sada: A polyherbal unani formulation. *J. Ethnopharmacol.* 2004; 91(2-3):219-223.
72. Halim ME, Misra A. The effects of the aqueous extract of *Pterocarpus santalinus* heartwood and vitamin E supplementation in streptozotocin-induced diabetic rats. *J. Med. Plants Res.* 2011; 5(3):398-409.
73. Yokozawa T, He LQ, Muto Y, Nagasaki R, Hattori M, Oura H. Effects of rhubarb extract in rats with diabetic nephropathy. *Phytother. Res.* 1997; 11(1):73-75.
74. Lee SH, Kim YS, Lee SJ, Lee BC. The protective effect of *Salvia miltiorrhiza* in an animal model of early experimentally induced diabetic nephropathy. *J. Ethnopharmacol.* 2011; 137(3):1409-1414.
75. Gondwe M, Kamadyaapa DR, Tufts M, Chuturgoon AA, Musabayane CT. *Sclerocarya birrea* [(A. Rich.) Hochst.] [Anacardiaceae] stem-bark ethanolic extract (SBE) modulates blood glucose, glomerular filtration rate (GFR) and mean arterial blood pressure (MAP) of STZ-induced diabetic rats. *Phytomed.* 2008; 15(9):699-709.
76. Bhuvaneswari P, Krishnakumari S. Nephroprotective effects of ethanolic extract of *Sesamum indicum* seeds (Linn.) in streptozotocin induced diabetic male albino rats. *Int. J. Green Pharm.* 2012; 6(4):330-335.
77. Soto C, Perez J, Garcia V, Uria E, Vadillo M, Raya L. Effect of silymarin on kidneys of rats suffering from alloxan-induced diabetes mellitus. *Phytomed.* 2010; 17(14):1090-1094.
78. Vessal G, Akmal M, Najafi P, Moein MR, Saqheb MM. Silymarin and milk thistle extract may prevent the progression of diabetic nephropathy in streptozotocin-induced diabetic rats. *Renal Failure* 2010; 32(6):733-739.
79. Ghaisas MM, Navghare VV, Takawale AR, Zope VS, Phanse MA. Antidiabetic and nephroprotective effect of *Tectona grandis* L. in alloxan induced diabetes. *Ars Pharmaceut.* 2010; 51(4):195-206.
80. Rao NK, Nammi S. Antidiabetic and renoprotective effects of the chloroform extract of *Terminalia chebula* Retz. seeds in streptozotocin-induced diabetic rats. *BMC Complement. Altern. Med.* 2006; 6:17.
81. Sharma S, Anjaneyulu M, Kulkarni SK, Chopra K. Resveratrol, a Polyphenolic Phytoalexin, is also found to attenuate diabetic nephropathy in rats. *Pharmacol.* 2006; 76(2):69-75.
82. Selvam R, Kalaiselvi P, Govindaraj A, Bala Murugan V, Sathish Kumar AS. Effect of *A. lanata* leaf extract and VEDIUPPU CHUNNAM on the urinary risk factors of calcium oxalate urolithiasis during experimental hyperoxaluria. *Pharmacol. Res.* 2001; 43(1):89-93.
83. Soundararajan P, Mahesh R, Ramesh T, Begum VH. Effect of *Aerva lanata* on calcium oxalate urolithiasis in rats. *Ind. J. Exp. Biol.* 2006; 44(12):981-986.
84. Prasad KV, Bharathi K, Srinivasan KK. Evaluation of *Ammannia baccifera* Linn. for antiurolithic activity in albino rats. *Ind. J. Exp. Biol.* 1994; 32(5):311-313.
85. Christina AJ, Ashok K, Packialakshmi M, Tobin GC, Preethi J, Muruges N. Antilithiatic effect of *Asparagus racemosus* Willd on ethylene glycol-induced lithiasis in male Albino Wistar rats. *Methods Find. Exp. Clin. Pharmacol.* 2005; 27(9):633-638.
86. Yasui T, Fujita K, Sato M, Sugimoto M, Iguchi M, Nomura S, *et al.* The effect of takusha, a kampo medicine, on renal stone formation and osteopontin expression in a rat urolithiasis model. *Urol. Res.* 1999; 27(3):194-199.
87. Byahatti VV, Pai KV, D' Souza MG. Effect of phenolic compounds from *Bergenia ciliata* (Haw.) Sternb. leaves on experimental kidney stones. *Ancient Sci. Life* 2010; 30(1):14-17.
88. Prachi, Chauhan N, Kumar D, Kasana MS. Medicinal plants of Muzaffarnagar district used in treatment of urinary tract and kidney stones. *Indian J. Tradit. Know.* 2009; 8(2):191-195.
89. Touhami M, Laroubi A, Elhabazi K, Loubana F, Zrara I, Eljahiri Y, *et al.* Lemon juice has protective activity in a rat urolithiasis model. *BMC Urol.* 2007; 7:18.
90. Odvina CV. Comparative value of orange juice versus lemonade in reducing stone-forming risk. *Clin. J. Amer. Soc. Nephrol.* 2006; 1(6):1269-1274.
91. Araujo Viel T, Diogo Domingos C, da Silva Monteiro AP, Riggio Lima-Landman MT, Lapa AJ, Souccar C. Evaluation of the antiurolithiatic activity of the extract of *Costus spiralis* Roscoe in rats. *J. Ethnopharmacol.* 1999; 66(2):193-198.
92. Varalakshmi P, Shamila Y, Latha E. Effect of *Crataeva nurvala* in experimental urolithiasis. *J. Ethnopharmacol.* 1990; 28(3):313-321.
93. Christina AJ, Packia Lakshmi M, Nagarajan M, Kurian S. Modulatory effect of *Cyclea peltata* Lam. on stone formation induced by ethylene glycol treatment in rats. *Methods Find. Exp. Clin. Pharmacol.* 2002; 24(2):77-79.
94. Golshan A, Hayatdavoudi P, Hadjzadeh MA, Khajavi Rad A, Mohamadian Roshan N, Abbasnezhad A, *et al.* Kidney stone formation and antioxidant effects of *Cynodon dactylon* decoction in male Wistar rats. *Avicenna J. Phytomed.* 2017; 7(2):180-190.
95. Hirayama H, Wang Z, Nishi K, Ogawa A, Ishimatu T, Ueda S, *et al.* Effect of *Desmodium styracifolium* triterpenoid on calcium oxalate renal stones. *Br. J. Urol.* 1993; 71(2):143-147.
96. Atmani F, Farell G, Lieske JC. Extract from *Herniaria hirsuta* coats calcium oxalate monohydrate crystals and blocks their adhesion to renal epithelial cells. *J. Urol.* 2004; 172(4):1510-1514.
97. Betanabhatla KS, Christina AJM, Syama BM. Antilithiatic activity of *Hibiscus sabdariffa* Linn. on ethylene glycol induced lithiasis in rats. *Nat. Prod. Rad.* 2009; 8(1):43-47.
98. Das M, Malipreddi H. Antiurolithiatic activity of ethanol leaf extract of *Ipomoea eriocarpa* against ethylene glycol-induced urolithiasis in male Wistar rats. *Ind. J. Pharmacol.* 2016; 48(3):270-274.
99. Bahuguna Y, Singh M, Rawat M, Juyal V, Gupta V. Antilithiatic effect of flowers of *Jasminum auriculatum* Vahl. *Int. J. Green Pharm.* 2009; 3(2):155-158.
100. Joyamma V, Rao SG, Hrishikeshavan HJ, Aroor AR, Kulkarni DR. Biochemical mechanisms and effects of *Mimosa pudica* (Linn) on experimental urolithiasis in rats. *Ind. J. Exp. Biol.* 1990; 28(3):237-240.
101. Karadi RV, Gadge NB, Alagawadi KR, Savadi RV. Effect of *Moringa oleifera* Lam. root-wood on ethylene glycol induced urolithiasis in rats. *J. Ethnopharmacol.* 2006; 105(1-2):306-311.
102. Poonguzhali PK, Chegu H. The influence of banana stem extract on urinary risk factors for stones in normal and hyperoxaluric rats. *Br. J. Urol.* 1994; 74(1):23-25.
103. Hadjzadeh MA, Khoei A, Hadjzadeh Z, Parizady M. Ethanolic extract of *Nigella sativa* L. seeds on ethylene glycol-induced kidney calculi in rats. *Urol. J.* 2007; 4(2):86-90.

104. Ramesh K, Manohar S, Rajeshkumar S. Nephroprotective activity of ethanolic extract of *Orthosiphon stamineus* leaves on ethylene glycol induced urolithiasis in albino rats. *Int. J. PharmaTech Res.* 2014; 6(1):403-408.
105. Barros ME, Lima R, Mercuri LP, Matos JR, Schor N, Boim MA. Effect of extract of *Phyllanthus niruri* on crystal deposition in experimental urolithiasis. *Urol. Res.* 2006; 34(6):351-357.
106. Tugcu V, Kemahli E, Ozbek E, Arinci YV, Uhri M, Erturkuner P, *et al.* Protective effect of a potent antioxidant, pomegranate juice, in the kidney of rats with nephrolithiasis induced by ethylene glycol. *J. Endourol.* 2008; 22(12):2723-2731.
107. Ogawa Y, Takahashi S, Kitagawa R. Effect of herb medicines for urolithiasis on urinary oxalate excretion in rats. *Acta Urol. Japon.* 1983; 29(10):1269-1271.
108. Vargas SR, Perez GRM. Diuretic and urolithiatic activities of the aqueous extract of the fruit of *Randia echinocarpa* on rats. *J. Ethnopharmacol.* 2002; 83(1-2):145-147.
109. Vargas SR, Perez GRM, Perez GS, Zavala SMA, Perez GC. Antiuro lithiatic activity of *Raphanus sativus* aqueous extract on rats. *J. Ethnopharmacol.* 1999; 68(1-3):335-338.
110. Grases F, Masarova L, Costa-Bauza A, March JG, Prieto R, Tur JA. Effect of "Rosa Canina" infusion and magnesium on the urinary risk factors of calcium oxalate urolithiasis. *Planta Med.* 1992; 58(6):509-512.
111. Christina AJ, Priya Mole M, Moorthy P. Studies on the antilithic effect of *Rotula aquatica* lour in male Wistar rats. *Methods Find. Exp. Clin. Pharmacol.* 2002; 24(6):357-359.
112. Divakar K, Pawar AT, Chandrasekhar SB, Dighe SB, Divakar G. Protective effect of the hydro-alcoholic extract of *Rubia cordifolia* roots against ethylene glycol induced urolithiasis in rats. *Food Chem. Toxicol.* 2010; 48(4):1013-1018.
113. Schneider HJ, Unger G, Rossler D, Bothor C, Berg W, Ernst G. Effect of drugs used for the prevention of urinary calculi recurrence on the growth and metabolism of young experimental animals. *Zeit. Urol. Nephrol.* 1979; 72(4):237-247.
114. Doddola S, Pasupulati H, Koganti B, Prasad KV. Evaluation of *Sesbania grandiflora* for antiuro lithiatic and antioxidant properties. *J. Nat. Med.* 2008; 62(3):300-307.
115. Jouad H, Lacaille-Duboi MA, Eddouks M. Chronic diuretic effect of water extracts of *Spergularia purpurea* in normal rats. *J. Ethnopharmacol.* 2001; 75(2-3):219-223.
116. Anand R, Patnaik GK, Kulshreshtha DK, Dhawan BN. Activity of certain fractions of *Tribulus terrestris* fruits against experimentally induced urolithiasis in rats. *Ind. J. Exp. Biol.* 1994; 32(8):548-552.
117. Laroubi A, Touhami M, Farouk L, Zrara I, Aboufatima R, Benharref A, *et al.* Prophylaxis effect of *Trigonella foenum graecum* L. seeds on renal stone formation in rats. *Phytother. Res.* 2007; 21(10):921-925.
118. Grases F, March JG, Ramis M, Costa-Bauza A. The influence of *Zea mays* on urinary risk factors for kidney stones in rats. *Phytother. Res.* 1993; 7(2):146-149.

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